

Short Note

# 1-(3-Amino-1-phenylpropyl)-3-(2-fluorophenyl)-1,3-dihydro-2*H*-benzimidazol-2-one

Catharina Neudorfer <sup>1,2,\*</sup>, Nadine Eberherr <sup>1</sup>, Karem Shanab <sup>1,2</sup>, Wolfgang Holzer <sup>2</sup>, Christina Rami-Mark <sup>1</sup>, Markus Mitterhauser <sup>1</sup>, Wolfgang Wadsak <sup>1</sup> and Helmut Spreitzer <sup>2,\*</sup>

- <sup>1</sup> Department of Biomedical Imaging and Image-guided Therapy, Division of Nuclear Medicine, Medical University of Vienna, Waehringer Guertel 18-20, 1090 Vienna, Austria; E-Mails: nadine\_e@gmx.at (N.E.); karem.shanab@gmail.com (K.S.); christina.rami-mark@meduniwien.ac.at (C.R.-M.); markus.mitterhauser@meduniwien.ac.at (M.M.); wolfgang.wadsak@meduniwien.ac.at (W.W.)
- <sup>2</sup> Division of Drug Synthesis, Department of Pharmaceutical Chemistry, Faculty of Life Sciences, University of Vienna, Althanstraße 14, 1090 Vienna, Austria; E-Mail: wolfgang.holzer@univie.ac.at
- \* Authors to whom correspondence should be addressed;
  E-Mails: catharina.neudorfer@gmail.com (C.N.); helmut.spreitzer@univie.ac.at (H.S.);
  Tel.: +43-4277-55629 (C.N.); +43-4277-55621 (H.S.); Fax: +43-4277-855629 (C.N.);
  +43-4277-855621 (H.S.).

Academic Editor: Norbert Haider

Received: 18 June 2015 / Accepted: 18 August 2015 / Published: 21 August 2015

Abstract: Starting from 1-(2-fluorophenyl)-1,3-dihydro-2H-benzimidazol-2-one (1) and (1-bromo-3-chloropropyl)benzene (2), the target compound 3, which represents a precursor for future radiolabeling, is prepared in a three-step synthesis.

Keywords: NET, PET, FAPPI

# 1. Introduction

The norepinephrine transporter (NET) plays a pivotal role in a variety of diseases, which not only include neurological/psychiatric disorders [1,2], but also cardiovascular [1–3] and metabolic diseases [3–5].

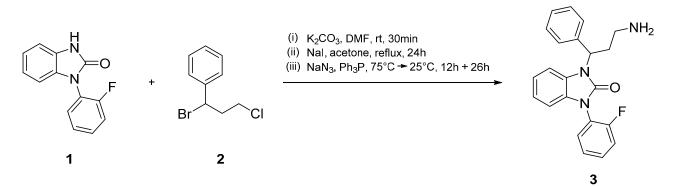
Thus, the investigation of the underlying dysregulation-mechanism of the noradrenergic system is of major interest.

As a non-invasive molecular imaging technique, positron emission tomography (PET) is the most suitable technique today to gain information about the transporter abundance and density in healthy and pathological living human brains [6]. It represents an accurate approach towards the collection of missing data in the living organism and also enables direct quantification of receptor and transporter densities *in vivo*. To fully gain insight into the molecular changes of the noradrenergic system via PET, however, prior development of suitable NET-PET radioligands is required. Currently, only PET tracers derived from reboxetine have been used in clinical studies [7] displaying certain limitations in their applicability, e.g. due to metabolic considerations [8].

On the basis of <sup>11</sup>C-radiolabeled 1-(3-(methylamino)-1-phenylpropyl)-3-phenyl-1,3-dihydro-2*H*-benzimidazole-2-one ([<sup>11</sup>C]Me@APPI) [9], which represents a suitable NET radioligand for use in PET, recently several non-radiolabeled analogs of [<sup>11</sup>C]Me@APPI have been described as potential reference compounds for PET based investigations of the NET [10]. In continuation of these previous studies, we are reporting in this paper the synthesis of 1-(3-amino-1-phenylpropyl)-3-(2-fluorophenyl)-1,3-dihydro-2*H*-benzimidazol-2-one (**3**), a precursor molecule for future radiolabeling operations.

#### 2. Results and Discussion

As previously published, derivative 1 was made accessible by reacting 1-fluoro-2-nitrobenzene with 2-fluoroaniline, followed by reduction of the nitro group. Subsequent cyclization with 1,1'-carbonyldiimidazole then afforded 1 [10]. For the preparation of side chain 2, the keto moiety of commercially available 3-chloro-1-phenylpropan-1-one was reduced with sodium borohydride and bromination of the resulting intermediate with aqueous hydrogen bromide led to the formation of 2 [10].



**Scheme 1.** Reaction of 1-(2-fluorophenyl)-1,3-dihydro-2*H*-benzimidazol-2-one (1) and (1-bromo-3-chloropropyl)benzene (2) to 1-(3-amino-1-phenylpropyl)-3-(2-fluorophenyl)-1,3-dihydro-2*H*-benzimidazol-2-one (3).

In the next reaction step, core compound 1 and side chain 2 were subjected to a condensation reaction under alkaline conditions (Scheme 1, (i)). Then, the chloro group of the resulting product was converted into an iodo group in a Finkelstein reaction (Scheme 1, (ii)). After purification, this intermediate was heated for 12 h in a solution of NaN<sub>3</sub> and DMF to obtain the respective azide

compound, which after treatment with triphenylphosphine eventually afforded target compound **3** (Scheme 1, (iii)).

### 3. Experimental Section

#### 3.1. General Information

The NMR spectrum was recorded from a CDCl<sub>3</sub> solution on a Bruker Avance III 400 spectrometer (Karlsruhe, Germany, 400 MHz for <sup>1</sup>H, 100 MHz for <sup>13</sup>C, 40 MHz for <sup>15</sup>N, 376 MHz for <sup>19</sup>F) at 25 °C. The center of the solvent (residual) signal was used as an internal standard which was related to trimethylsilane (TMS) with  $\delta$  7.26 ppm (<sup>1</sup>H in CDCl<sub>3</sub>), and  $\delta$  77.0 ppm (<sup>13</sup>C in CDCl<sub>3</sub>). <sup>19</sup>F-NMR spectra were referenced by absolute referencing via  $\Xi$  ratio. Digital resolutions were 0.25 Hz/data point in the <sup>1</sup>H and 0.3 Hz/data point in the <sup>13</sup>C-NMR spectra. Coupling constants (*J*) are quoted in Hz. The following abbreviations were used to show the multiplicities: s: singlet, d: doublet, t: triplet, q: quadruplet, dd: doublet of doublet, m: multiplet. Mass spectra were obtained on a Shimadzu QP 1000 instrument (Kyoto, Japan, EI, 70 eV), high-resolution mass spectrometry (HRMS) was carried out on a maXis HD ESI-Qq-TOF mass spectrometer (Bruker Daltonics, Bremen, Germany) in the positive-ion mode by direct infusion. Compound purity: all compounds synthesized featured a purity of at least 95%.

# 3.2. 1-(3-Amino-1-phenylpropyl)-3-(2-fluorophenyl)-1,3-dihydro-2H-benzimidazol-2-one

(i) To a solution of **1** (1.00 g, 4.38 mmol) in DMF (5 mL) was added K<sub>2</sub>CO<sub>3</sub> (1.21 g, 8.76 mmol) and the resulting mixture was stirred for 30 min at 25 °C. After 30 min, **2** (1.53 g, 6.57 mmol) was added and stirring was continued overnight. Ethyl acetate (5 mL) and water (5 mL) were added to the mixture and the aqueous layer was extracted several times with ethyl acetate (10 mL). The combined organic layers were washed with brine, dried over MgSO<sub>4</sub>, and evaporated to dryness. Purification of the resulting product was carried out by column chromatography (silica gel 60) with petroleum ether/ethyl acetate 9:1 to afford an orange resin.

(ii) The resulting orange resin (0.84 g, 2.20 mmol) was dissolved in acetone (7 mL) and NaI (0.66 g, 4.40 mmol) was added. The mixture was refluxed for 24 h, after which the formed precipitate was filtered and concentrated prior to purification via column chromatography (silica gel 60) with petroleum ether/ethyl acetate 9:1, to give the intermediate product as yellow crystals.

(iii) The obtained product (0.15 g, 0.32 mmol) was dissolved in DMF (3 mL) and heated to 75 °C upon addition of NaN<sub>3</sub> (0.05 g, 0.65 mmol). After 12 h the reaction was quenched with H<sub>2</sub>O (10 mL) and the resulting mixture was extracted with ethyl acetate ( $3 \times 30$  mL). The organic layer was washed with brine ( $3 \times 10$  mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure. The crude product was purified via column chromatography (silica gel 60, petroleum ether/ethyl acetate 8.5:1.5) and was introduced in the subsequent reaction step.

Triphenylphosphine (0.07 g, 0.27 mmol) was added to a solution of the prior synthesized azide (0.07 g, 0.18 mmol) in THF (3 mL) and the mixture was stirred for 10 h at 25 °C. Thereafter, H<sub>2</sub>O was added and the reaction was stirred for another 16 h. The resulting reaction product was concentrated *in vacuo* and purified by column chromatography (silica gel 60, CH<sub>2</sub>Cl<sub>2</sub>/MeOH 9:1).

Yield: 0.04 g (57%), colorless crystals, m.p. 135-136 °C.

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) 2.44 (br s, 2H, NH<sub>2</sub>), 2.50–2.54 (m, 2H, 2'-C<u>H<sub>2</sub></u>), 2.73–2.84 (m, 2H, 3'-C<u>H<sub>2</sub></u>), 5.88–5.90 (m, 1H, 1'-C<u>H</u>), 6.82–6.84 (m, 2H, benzim 7-C<u>H</u>, benzim 4-C<u>H</u>), 6.93–7.02 (m, 2H, benzim 6-C<u>H</u>, benzim 5-C<u>H</u>), 7.26–7.37 (m, 5H, phen 4-C<u>H</u>, f-phen 3-C<u>H</u>, f-phen 5-C<u>H</u>), 7.42–7.57 (m, 4H, f-phen 6-C<u>H</u>, f-phen 4-C<u>H</u>, phen 2-C<u>H</u>, phen 6-C<u>H</u>).

<sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  (ppm) 33.6 (2'-<u>C</u>H<sub>2</sub>), 38.6 (3'-<u>C</u>H<sub>2</sub>), 53.1 (1'-<u>C</u>H), 108.8 (d, J = 1.4 Hz, benzim 4-<u>C</u>H), 109.9 (benzim 7-<u>C</u>H), 117.1 (d, J = 19.5 Hz, f-phen 3-<u>C</u>H), 121.4 (benzim 5-<u>C</u>H), 121.9 (benzim 6-<u>C</u>H), 122.0 (f-phen 1-<u>C</u>H), 124.9 (d, J = 3.9 Hz, f-phen 5-<u>C</u>H), 127.2 (phen 2-<u>C</u>H), 127.2 (phen 6-<u>C</u>H), 127.7 (phen 4-<u>C</u>H), 128.1 (benzim 7a-C), 128.7 (phen 3-<u>C</u>H), 128.7 (phen 5-<u>C</u>H), 129.6 (f-phen 6-<u>C</u>H), 129.6 (benzim 3a-C), 130.2 (d, J = 7.8 Hz, f-phen 4-<u>C</u>H), 138.7 (phen 1-C), 153.5 (benzim 2-<u>C</u>O), 157.9 (d, J = 253.0 Hz, f-phen 2-<u>C</u>F).

<sup>19</sup>F-NMR (471 MHz, CDCl<sub>3</sub>): δ (ppm) –118.58 (m, f-phen C<u>F</u>), MS: *m/z* (%) 361 (M<sup>+</sup>, 17), 331 (10), 318 (11), 228 (100), 199 (21), 185 (18), 133 (20), 103 (17), 91 (32), 77 (25), 43 (25).

HRMS: *m/z* calculated for C<sub>22</sub>H<sub>21</sub>FN<sub>3</sub>O [M + H]<sup>+</sup>: 362.1663. Found: 362.1665.

# **Author Contributions**

Catharina Neudorfer: Responsible for the performance of the syntheses, supervision and writing; Nadine Eberherr: Performance of the syntheses; Karem Shanab: Contributions to syntheses and experimental procedures; Wolfgang Holzer: Performance of the NMR analyses; Christina Rami-Mark: Designed parts of the research; Markus Mitterhauser: Designed parts of the research and proofread the manuscript; Wolfgang Wadsak: Designed parts of the research and proofread the manuscript; Helmut Spreitzer: Conceived and supervised the syntheses.

# **Conflicts of Interest**

The authors declare no conflict of interest.

# References

- Sung, U.; Apparsundram, S.; Galli, A.; Kahlig, K.M.; Savchenko, V.; Schroeter, S.; Quick, M.W.; Blakely, R.D. A regulated interaction of syntaxin 1A with the antidepressant-sensitive norepinephrine transporter establishes catecholamine clearance capacity. *J. Neurosci.* 2003, 23, 1697–1709.
- Kim, C.H.; Hahn, M.K.; Joung, Y.; Anderson, S.L.; Steele, A.H.; Mazei-Robinson, M.S.; Gizer, I.; Teicher, M.H.; Cohen, B.M.; Robertson, D.; *et al.* A polymorphism in the norepinephrine transporter gene alters promoter activity and is associated with attention-deficit hyperactivity disorder. *Proc. Natl. Acad. Sci. USA* 2006, *103*, 19164–19169.
- 3. Hahn, M.K.; Robertson, D.; Blakely, R.D. A mutation in the human norepinephrine transporter gene (SLC6A2) associated with orthostatic intolerance disrupts surface expression of mutant and wild-type transporters. *J. Neurosci.* **2003**, *23*, 4470–4478.

- Mirbolooki, M.R.; Upadhyay, S.K.; Constantinescu, C.C.; Pan, M.L.; Mukherjee, J. Adrenergic pathway activation enhances brown adipose tissue metabolism: A [<sup>18</sup>F]FDG PET/CT study in mice. *Nucl. Med. Biol.* 2014, 41, 10–16.
- Lin, S.L.; Fan, X.; Yeckel, C.W.; Weinzimmer, D.; Mulnix, T.; Gallezot, J.D.; Carson, R.E.; Sherwin, R.S.; Ding, Y.S. *Ex vivo* and *in vivo* Evaluation of the Norepinephrine Transporter Ligand [<sup>11</sup>C]MRB for Brown Adipose Tissue Imaging. *Nucl. Med. Biol.* 2012, *39*, 1081–1086.
- 6. Wadsak, W.; Mitterhauser, M. Basics and principles of radiopharmaceuticals for PET/CT. *Eur. J. Radiol.* **2010**, *73*, 461–469.
- Rami-Mark, C; Zhang, M.R.; Mitterhauser, M.; Lanzenberger, R.; Hacker, M.; Wadsak, W. [18F]FMeNER-D2: Reliable fully-automated synthesis for visualization of the norepinephrine transporter. *Nucl. Med. Biol.* 2013, 40, 1049–1054.
- Vanicek, T.; Spies, M.; Rami-Mark, C.; Savli, M.; Höflich, A.; Kranz, G.S.; Hahn, A.; Kutzelnigg, A.; Traub-Weidinger, T.; Mitterhauser, M.; *et al.* The norepinephrine transporter in attention-deficit/hyperactivity disorder investigated with positron emission tomography. *JAMA Psychiatry* 2014, *71*, 1340–1349.
- Mark, C.; Bornatowicz, B.; Mitterhauser, M.; Hendl, M.; Nics, L.; Haeusler, D.; Lanzenberger, R.; Berger, M.L.; Spreitzer, H.; Wadsak, W. Development and automation of a novel NET-PET tracer: [C-11]Me@APPI. *Nucl. Med. Biol.* 2013, 40, 295–303.
- Neudorfer, C.; Seddik, A.; Shanab, K.; Jurik, A.; Rami-Mark, C.; Holzer, W.; Ecker, G.; Mitterhauser, M.; Wadsak, W.; Spreitzer, H. Synthesis and *in Silico* Evaluation of Novel Compounds for PET-Based Investigations of the Norepinephrine Transporter. *Molecules* 2015, 20, 1712–1730.

© 2015 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/4.0/).